

Masood Sadiq Butt
Muhammad Tahir-Nadeem
Muhammad Kashif Iqbal Khan
Rabia Shabir
Mehmood S. Butt

Oat: unique among the cereals

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M. Sadiq Butt · M. Tahir-Nadeem (✉)
M.K.I. Khan · R. Shabir
Institute of Food Science and Technology
University of Agriculture
Faisalabad, Pakistan
E-Mail: tahir_nadeem@yahoo.com

M.S. Butt
Medical Directorate
Withybush General Hospital
Wales, UK

■ **Abstract** This review is intended to focus on the composition of oat and its therapeutic potential in the pharmacology that supports its use to cure various maladies. Oat (*Avena sativa*) is distinct among the cereals due to its multifunctional characteristics and nutritional profile. Recent advancement in food and nutrition has revealed the importance of its various components. It is a good source of dietary fiber especially β -glucan, minerals and other nutrients. Oat and oat by products have been proven to be helpful in the treatment of diabetes and cardiovascular disorders. Oat bran in particular, is good source of B complex vitamins, protein, fat, minerals besides heart healthy soluble fiber β -glucan. The β -glucan has outstanding functional properties and is of immense importance in human nutrition.

Different physiological effects of β -glucan are related to its viscosity, attenuation of postprandial plasma glucose and insulin responses, high transport of bile acids towards lower parts of the intestinal tract and high excretion of bile acids thereby lowering of serum cholesterol levels. Moreover, it is helpful against coeliac disease. The incorporation of oat grains and oat bran in the food products improves not only the nutrition but also a therapy against various maladies.

■ **Key words** oat – oat bran – β -glucan – dietary fiber – cardiovascular diseases – hyperglycemia

Introduction

Oat, like all other grain varieties, belongs to the Poaceae family and is known as “Jai” or “Javi” in Indian subcontinent. *Avena sativa* L (common oat) is the most important among the cultivated oats (Table 1). Like wheat, it is an annual grass that is believed to be Asiatic in origin. During early growth, the oat plant consists of leaves and a shortened stem, giving a rosette type of plant. The tillers grow into additional

“branch plants” or tillers and under favorable conditions, the plant can form up to 30 tillers. The main stem and tillers can reach up to 2 or more ft, depending on variety and growing condition. These stems terminate in a large panicle that bears flowers and seeds or kernels. Each main and lateral stem as well as branch stem terminates in a spikelet that is removed during threshing. Generally two kernels, but occasionally one, are produced per spikelet. The oat kernel, also termed caryopsis or groat, is the part after the removal of palea and lemma. It is elon-

Table 1 Taxonomic information

Botanical name	<i>Avena sativa</i>
Kingdom	Plantae: plants
Subkingdom	Tracheobionta: vascular plants
Superdivision	Spermatophyta: seed plants
Division	Magnoliophyta: flowering plants
Class	Liliopsida: monocotyledons
Subclass	Commelinidae
Order	Cyperales
Family	Poaceae: grass family
Genus	<i>Avena</i> : oat
Species	<i>A. sativa</i> : common oat, <i>A. byzantina</i> , <i>A. fatua</i> , <i>A. diffusa</i> , <i>A. orientalis</i>

gated—spindle shaped, up to about 0.5 in. in length and 0.125 in. or less in width. It is generally covered with fine, silky hairs and includes the seed coat layers of cells, starchy endosperm and the embryo. Oat is an important food grain in temperate regions of the world. Modern oat probably originated from the Asian wild red oat, which grew as a weed in other grain crops [76].

It is an annual crop used both for human and animal nutrition. Before being used as a food, it was used for medicinal purposes. With the development in field of nutrition, oat was recognized as a healthy food in the mid 1980s signifying that a substance in it helped prevent heart disease and therefore it became more popular for human nutrition [114]. Since oats are not suitable for bread making due to lack of gluten therefore often served as a porridge, flakes or breakfast cereals made from crushed or rolled oats. As oat flour or oatmeal, it is utilized in a variety of baked items like composite bread made from mixture of oatmeal and wheat flour. When the objective is to utilize oat flour, millers are more conscious about its protein contents. While oat cultivars are characterized as to their relative protein concentration, little is known about genetic differences in elemental composition, which may also be of nutritional significance for food and feed. Oat has a couple of traits that causes it to be less favored than other grains—a bland taste and a tendency to spoil. Despite these issues, oat is a staple in Germany, Ireland, Scotland, and the Scandinavian countries [102].

Grain

Oat grain has soft kernel and lipid distributed throughout the seed, which makes its milling process more difficult than wheat and corn. To prevent from atmospheric oxidation, the oat is given a hydrothermic treatment before processing. Hull (husk) of oat grain is about 25–30% of the seed (Fig. 1)

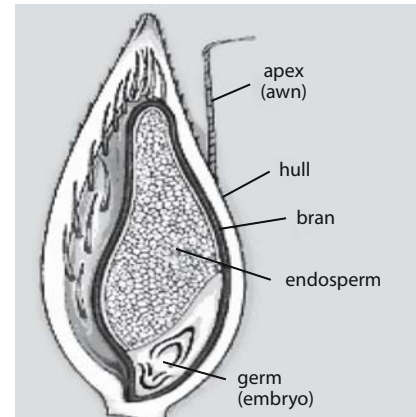


Fig. 1 Cross section of oat grain. By courtesy of Encyclopedia Britannica Inc., copyright 1996; used with permission

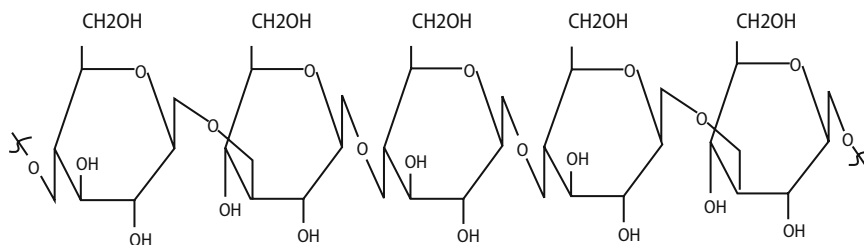
The grain is dehulled before use, whereas husk after processing may be used in food industry. Unprocessed hull contain silicate particles, which have barbed nature and can irritate the mouth, esophagus and gastrointestinal tract. Whole oat groat contains high amounts of valuable nutrients such as soluble fibers, proteins, unsaturated fatty acids, vitamins, minerals, and phytochemicals (Table 2). The dietary fiber complex with its antioxidants and other phytochemicals is effective against cardiovascular disease and some types of cancer [49, 50, 51, 101, 108].

Oat groat contains significant amounts of β -glucan that varies between 2.3 and 8.5 g/100 g [113]. β -glucan in oat is distributed through the endosperm and is located in the endosperm cell walls constituting about 75% of the endosperm cell walls. It is also present in the aleurone cell wall lesser than in endosperm [80].

Table 2 Oat grain composition

Component	% Age
Moisture	13.3
Protein	13
Lipids	7.5
Fiber	10.3
Ash	3.1
Calcium (mg/100 g)	60
Phosphorus (mg/100 g)	372
Iron (mg/100 g)	3.8
Zinc (mg/100 g)	3.9
Iodine (mg/100 g)	16
Thiamin (mg/100 g)	0.50
Riboflavin (mg/100 g)	0.14
Niacin (mg/100 g)	1.3
Energy (MJ/100 g)	1.61

Kirk and Sawyer [71]

Fig. 2 β -glucan

■ Bran

Bran is the edible, outermost layer of the oat kernel and is produced by grinding clean groats or rolled oats for separating the resulting flour by sieving, bolting and other suitable means into fraction such that the oat bran is not more than 50% of the starting material. It has total β -glucan and dietary fiber not less than 5.5 and 16.0% respectively with at least one third of total dietary fiber is soluble fiber [8]. Like oatmeal, oat bran contains B complex vitamins, protein, fat, minerals, and heart healthy soluble fiber called β -glucan. Oat bran contains 17.1% protein, 67.9% carbohydrates, 8.6% fat, 15–22% dietary fiber, 10.4% β -glucan, 1.3 mg niacin, 171 mg magnesium, 6.4 mg iron, 0.17 mg copper, 441 mg potassium and α -tocopherol less than 0.5 mg [78, 98].

■ Dietary fiber

Dietary fiber is found only in plant foods. It is the remnant of the edible part of plants that are resistant to human digestion system [1]. It consists of both soluble and insoluble fiber. Soluble fiber dissolves while insoluble fiber does not dissolve in water. Both types are important to health in different ways. Soluble fiber includes gums, mucilages, pectin and some hemicelluloses. Cellulose, lignin (polymerized *p*-coumaryl alcohols) and the rest of the hemicelluloses, are all insoluble fibers. Water-soluble fiber in cereals is composed of non-starchy polysaccharides such as β -glucan. Water-soluble dietary fiber can form viscous solutions. Increased viscosity in the intestine slows intestinal transit, delays gastric emptying [2, 116], and slows glucose and sterol absorption in the intestine. The soluble fiber from oatmeal and oat bran is very effective to lower blood cholesterol and normalize blood sugar levels [58, 118, 120]. Insoluble fiber contains lignin as well as non-starchy polysaccharides. Lignin is not a polysaccharide but is a lipophilic, phenolic polymer, which can absorb bile acids. Insoluble dietary fibers usually have high water-holding capacity, which contributes to increased fecal bulk.

Associated with dietary fiber are antinutrients, such as phytic acid and oxalic acid, and proteins which could affect to a certain extent mineral bio-availability by binding and trapping minerals within dietary fiber particles or shortening the transit time of nutrients through the intestine. There are several *in vivo* and *in vitro* studies which indicate that dietary fiber might have important impacts on mineral balance [45, 48].

■ β -glucan

Dietary fiber has important component β -D-glucan. Purified oat β -glucan is linear, unbranched polysaccharide composed of 1-4-*O*-linked (70%) and 1-3-*O*-linked (30%) β -D-glucopyranosyl units (Fig. 2). The 1-3-linkages occur singly and most of the 1-4-linkages occur in groups of two or three leading predominantly to a structure of β -(1-3)-linked cellotriosyl and cellotetraosyl units [28, 83, 118]. On the basis of linkage analysis oat β -glucan and non-cereal β -glucan appear identical but analysis of oligosaccharides fragments released by enzymatic hydrolysis reveals differences.

β -glucan has outstanding functional and nutritional properties exhibiting high viscosities at relatively low concentrations. β -glucan solutions (concentration: 1%) have a low flow behavior index and a high consistency index in the power law model [9]. Its viscosity is stable over a wide range of pH (2–10) but it decreased with increasing temperature [29] Because of its viscosity, β -glucan may disturb the brewing process or limit the nutritive value of oat-based feeds for animals [23]. On the other hand, β -glucan can be used as a thickening agent in food industry [117]; it may influence the sensory quality of beverages [75] and is of particular importance in human nutrition [82].

Different physiological effects of β -glucan in isolated form or as a constituent of oat and barley products are related to its viscosity: attenuation of postprandial plasma glucose and insulin responses [121], high transport of bile acids towards lower parts of the intestinal tract and high excretion of bile acids [74] or lowering of serum cholesterol levels [17, 79].

Molecular weight and concentration have a great influence on the viscosity and the rheological behavior of β -glucans in aqueous solution and in the intestinal tract. Difference in rheological properties of β -glucan from several oat varieties was found at the same β -glucan concentration due to difference in molecular weight [10].

β -Glucans extracted from oat bran has higher viscosities than those from oat endosperm [115]. Similarly β -glucans extracted from enhanced oat lines are more viscous than those from traditional lines [26]. In order to be physiologically active and form viscous solutions in the gut, β -glucan must be soluble, and the concentration and molecular weight must be sufficiently high and vary between 2.68×10^4 and 3×10^6 g/mol [3, 28, 35]. The molecular weight of β -glucan in oat/barley products is reported to be smaller than the molecular weight of β -glucan in the raw material [3, 13, 66, 106]. The molecular weight of β -glucan in oat bread, for example, was reduced as compared with the molecular weight of β -glucan in oat bran. Raw material, endogenous β -glucanase activity, processing, and storage conditions affect the amount, solubility, molecular weight, and structure of β -glucan in the products [13, 31, 123].

There are some indications that the molecular weight of β -glucan may partly be reduced during its passage through the upper gastrointestinal tract [56, 92]. However controversial reports are available that shows no quantitative losses of β -glucan from oat flour or bran in the stomach and in upper parts of the small intestine in ileum-cannulated pigs [11].

β -glucan containing extrudates from oat affect bile acid binding and fermentation in vitro [37]. Oat products of different composition, e.g. oat meal, oat bran etc. pre-treated by different methods, e.g. extrusion, autoclavation or even untreated when used in diets, have shown beneficial effects in nutritional studies with humans and rats [36]. During passage through the stomach and small intestine, insoluble β -glucans are partly converted into soluble form, and can form viscous solutions in the gut. There is only limited information on the flow behavior of β -glucans from different oat sources used in nutrition [34].

■ Oat and diabetes

Oat contains β -glucan that controls blood glucose and cardiovascular diseases. Different researchers have investigated the effects of high β -glucan oat bran flour on patients with Type II diabetes [54, 107]. Twelve patients, for example, were administered oat bran flour, oat bran crisp and a glucose load providing 12.5 g glycemic carbohydrate (Series 1), and 25 g

glucose load alone and 25 g glucose load with 30 g oat bran flour (Series 2). Study participants undertook five 2-h meal glucose tolerance tests on separate occasions. Oat bran flour high in β -glucan had a low glycemic response and decreased postprandial glycemic response of an oral glucose load in subjects. Canadian researchers similarly evaluated the long-term effects of oat bran concentrate on free-living subjects with Type II diabetes, in which eight subjects were fed bread containing oat bran concentrate (soluble fiber β -glucan content = 22.8%), followed by a white bread control in the second phase of the trial [55].

Test subjects' mean total plasma cholesterol and LDL cholesterol levels were lower in the oat bran concentrate period than in the white bread period of the study, and the mean ratio of LDL cholesterol to HDL cholesterol was reduced by 24% in the oat bran concentrate phase of the trial. The oat bran concentrate bread products improved lipidemic as well as glycemic and insulinemic responses through its ability to lower total and LDL cholesterol. Consumption of oat extracts containing soluble glucans causes decline in glucose, insulin and glucagon responses to carbohydrates loads. Incorporation of very modest amounts of the soluble oat extract (50–75 g/day) into foods affects risk factors for disease without altering the acceptability or palatability of diets [57]. While comparing acute responses of healthy subjects to glucose or foods containing glucose plus oat gum, the oat gum shows more palatability [17].

Normally glucagon value is expected to decline after a carbohydrate load; however, in patients with non-insulin dependent diabetes, glucagon secretion has been reported to increase inappropriately after a carbohydrate meal. Although subjects in the study were not diabetic, hyperinsulinemia suggests an abnormality of glucose metabolism, which is supported by the increased glucagon concentrations after the carbohydrate load. The comparative reduction of glucagon response after the oat extracts may be partially responsible for the reduction in glucose concentrations [57].

■ Cholesterol and cardiovascular diseases

Coronary artery disease is the major cause of death in the United States and in most Western countries and blood cholesterol is a major risk factor [64]. Dietary and pharmacologic reductions in total and LDL cholesterol decrease the risk of the malady [22, 94], and dietary intervention is the first-line approach. Increasing dietary fiber has been recommended as a safe and practical approach for cholesterol reduction [110]. On the basis of numer-

ous clinical studies, the US Food and Drug Administration (FDA) permitted the use of a claim that oat-soluble fiber has the ability to reduce the risk of heart disease. The required dose of β -glucan for a single food is 0.75 g/serving. In literature, the highly viscous β -glucan fraction of oat has been reported to lower blood cholesterol and the intestinal absorption of glucose [81, 119]. Similar behavior has been observed for other soluble fibers such as psyllium, pectin, and guar gum [41, 111].

Oat bran exerts a small but potentially useful effect on plasma lipoprotein risk factors for cardiovascular disease. In subjects with mild hypercholesterolemia and normal blood pressure when receive different diets containing dietary fiber from wheat, oat, and rice bran; the oat bran has been found to be the only fiber source that significantly lowered total and low-density-lipoprotein (LDL) cholesterol levels. Although all three brans were found to slightly increase high-density-lipoprotein (HDL) cholesterol levels there were no significant changes in blood pressure, blood glucose, or serum insulin responses to a test meal on any of the bran-supplemented diets. However, this potential benefit may be limited by the amount of fiber that individuals would need to consume [67].

Water insoluble wheat fiber and cellulose have no effect on the cholesterol levels unless they displace foods supplying saturated fats and cholesterol. Although soluble fibers are more effective in lowering blood cholesterol levels, there is debate as to the degree of cholesterol reduction caused by them. The cholesterol lowering potential varies extensively among different fiber sources. The reasons for such ample variations include small sample sizes, different dosages of fiber, different background diets, concurrent changes in body weight, varying dietary control, and different types of subjects. Certain fibers lower cholesterol more effectively than others and oat is an example in this regard [4, 14, 72, 91].

Concurrent changes in fat and cholesterol caused by inadequate dietary control can confound the relation between increased fiber intake and blood cholesterol concentrations. For this reason, quantitating the direct effect of soluble fiber on cholesterol lowering, in addition to that attributed to displacement of saturated and trans-unsaturated fat in the diet, is difficult. Oat bran due to high content of soluble fiber is recommended as a potentially beneficial adjunct to lipid-lowering diets [99]. This recommendation is supported by the results of studies that demonstrate significant, although variable, reductions in serum cholesterol after ingestion of various oat bran-containing products [6, 70, 91].

Kahlon et al. [62] reported significant plasma cholesterol reductions with rice bran and oat bran

diets in growing hamsters fed 10% fat and 0.5% cholesterol diets for 3 weeks. LDL values in hamsters fed cellulose + soy protein + vitamin E; Rice bran, OB, and oat bran + vitamin diets were significantly lower than in those fed the control diet, resulting in reduced atherogenic risk in those animals. Animals fed on oat bran and oat bran + vitamin E diets had significantly lower liver cholesterol concentrations when compared with all other groups, suggesting oat bran diets more effective to lower liver cholesterol accumulation as well as for reduced fatty infiltration of the liver. A mechanism for cholesterol reduction with oat bran diets can include an increased excretion of bile acid [30], which in turn stimulates liver to utilize available cholesterol to produce more bile acid [59, 60, 62, 63]. Intake of oat bran results in increased lipid excretion and percent digestibility. Lipid excretion may be different among various dietary fiber sources. Diets containing rice bran and oat bran significantly diminished the atherogenicity of 20% fat, 0.5% cholesterol diets in hamsters as manifested by a 45–65% reduction in foam cell formation in the aortic arch of the animals. The effect is apparently influenced by reduction in the plasma LDL level and LDL / HDL ratios [61].

Cereal-fiber sources decrease the chylomicron cholesterol concentration. The postprandial serum triglyceride response is significantly lowered with the addition of oat bran. Several mechanisms can be involved, such as impaired dietary triglyceride absorption from the small intestine, increased clearance of chylomicron particles and chylomicron remnant uptake, reduced liver very-low-density lipoprotein (VLDL) secretion, or increased VLDL catabolic rate [77].

If some food source causes interruption in the synthesis of cholesterol by the body it can lower cholesterolemia. Foods containing high doses of cholesterol can decrease cholesterolemia for several hours as compared with the fasting value [24]. Several complementary mechanisms are likely to be involved. First, a single amount of cholesterol from intestinal origin, on entering the blood stream causes the liver to quickly reduce the endogenous synthesis of cholesterol. This lowers postprandial cholesterolemia early after the meal intake. This effect results from the inhibition of the activity of key enzyme involved in cholesterol synthesis, HMG-CoA reductase, as shown in laboratory animals [21]. Fiber sources are able to decrease the chylomicron cholesterol secretion and a marked reduction of cholesterolemia is observed.

The lowering effect of rice bran on chylomicron cholesterol (22.2%) has been found not to be sufficient to significantly lower postprandial cholesterolemia whereas oat bran, which decreased chylomicron

cholesterol by 43.3%, significantly reduced postprandial cholesterolemia. Oat bran containing high amounts of soluble fibers which repeatedly displayed a hypo cholesterolemia effect after chronic intake [7] exhibited the most marked influence on postprandial cholesterolemia. Another mechanism possibly involved in the reduction of postprandial cholesterolemia can be an inhibition of the HMG-CoA reductase activity by fiber sources. This has been shown to occur with various cereal fractions in animal models [86]. In the presence of oat bran, the mean area of variation of serum cholesterol minus chylomicron cholesterol (-2.67 mmol/L h) has been found significantly lower than that given by the control test meal (-1.71 mmol/L h).

■ Oat and lipids

Dietary fiber reduces fasting lipoproteins and postprandial lipoproteins [5, 87]. Oat bran significantly decreases serum total cholesterol and LDL cholesterol compared with control values. Anderson et al. [5] compared the effects of soluble-fiber (oat bran) and insoluble fiber (wheat bran) intakes on serum lipids, apolipoproteins, and lipoprotein fractions for 20 hypercholesterolemic men on a metabolic ward for 21 day. They postulated that the interactions of four separate processes may contribute to the hypocholesterolemic effect of oat bran. First, oat bran significantly increases fecal bile acid excretion and alters bile acid metabolism. Second, oat bran may alter lipoprotein metabolism, possibly by increasing hepatic LDL receptors. Oat bran tends to selectively lower LDL cholesterol to a greater extent than does HDL cholesterol. Third, oat bran is fermented in the colon into short chain fatty acids such as acetate, propionate, and butyrate and after absorption into the portal vein, propionate may inhibit hepatic cholesterol synthesis [122]. Fourth, decreases in insulin secretion associated with fiber such as oat bran could lead to reduction in cholesterol synthesis [55]. A daily intake of 106 g oat bran providing 15.3 g TDF (total dietary fiber) and 7.6 g soluble-fiber over 21 day was accompanied by significant reduction in total cholesterol, LDL cholesterol, and apolipoprotein B-100 in hypercholesterolemic men. An equivalent intake of TDF from wheat bran providing 1.3 g soluble-fiber did not significantly reduce these contents.

In humans, acetate is an important large bowel digestion product of diets containing oat products. Susan et al. [18] compared the oat bran and wheat bran along with control for the production of serum acetate in human body. They got the serum acetate concentrations at 0800 hours ranged from 52 to

83 μ mol/L for all subjects that were similar to the values for fasting serum acetate in humans as reported by Guynn and Veech [44]. Fasting serum acetate values were significantly higher after the oat-bran than after the control diets. Serum acetate concentration was significantly higher in subjects fed oat bran than in subjects fed wheat bran. Serum acetate concentrations for the oat-bran group peaked at 1400 (109 ± 12 μ mol/L) and 1600 hours (113 ± 13 μ mol/L) [18].

Similar diet-induced differences in short chain fatty acids (SCFA) concentrations in both cecal contents and hepatic portal venous plasma have been noted previously in animals fed highly degradable fibers such as pectin, beans, and oat bran and marginally degradable fibers such as cellulose and wheat bran [39, 104]. It was reported that although acetate was the principal SCFA produced in rats by oat bran and cellulose diets, propionate was significantly higher in the portal vein of rats fed oat-bran diets compared with cellulose diets [25].

Significantly higher concentration of SCFAs have been reported both in hepatic portal venous plasma and fecal contents of rats fed oat-bran diets compared with cellulose diets. Acetate was the major SCFA produced by both diets. However, the animals fed oat bran had proportionately less acetate and more propionate and butyrate than in animals fed cellulose. This finding may be due to the fermentation of complex carbohydrates, such as the hemi-cellulose present in oat bran, leading to fermentation of longer-chain acids [89].

Concentrations of SCFAs have been shown to decrease in the blood as they pass from the portal to hepatic to peripheral sites. At all sites acetate is the principal anion but molar ratios of the three principal SCFAs change at various sites, indicating greater uptake of butyrate by the colonic epithelium and of propionate by the liver [27]. Increases in peripheral serum acetate in humans, therefore, are likely accompanied by increases in portal vein acetate and propionate as well. Peripheral serum acetate was also noted to be significantly higher in oat-bran fed subjects than in wheat-bran fed subjects [7]. Both acetate [15] and propionate [122] have been reported to inhibit cholesterol synthesis in animals. Thus, significant increases in SCFAs in humans may contribute to the hypocholesterolemic effects of oat bran and other sources of soluble-fiber.

Oat bran is effective in reducing the risk for CHD because it favorably alters atherogenic fasting and postprandial serum lipids and other lipoprotein fractions, independent of other dietary changes. Carefully controlled studies of longer duration are needed to determine if the hypocholesterolemic effect of oat bran is sustained [5].

■ Oat and coeliac disease

Coeliac disease is an autoimmune hereditary disorder of the small intestine that occurs in people of all ages from middle infancy because of sensitivity to gluten in food. Normally the lining of the small intestine has a fluffy velvety texture, but in coeliac disorder it becomes smooth and flat. This reduces its ability to absorb nutrients, including sugars, proteins, vital minerals and vitamins from food. When persons with coeliac disease take foods containing gluten, their immune system responds by damaging the small intestine lining. Tiny fingerlike protrusions, called villi are attacked by the immune system and are eventually destroyed. Malnutrition occurs without these villi, no matter how much food a person consumes because the nutrients from food pass the gut without being absorbed (malabsorption), leading to diarrhoea, vitamin and mineral deficiencies, anaemia and osteoporosis. Children usually develop it at between six and 18 months of age. However, the onset of the disorder can be delayed and it can occur at any age, when the symptoms come on slowly, perhaps over years, making early diagnosis difficult [38, 100].

Presently, the only effective treatment of coeliac disease is a life-long rely on gluten-free diet [73]. No medication exists that will prevent damage, or keep the body from attacking the gut when gluten is present. Strict adherence to the gluten free diet allows the intestines to heal, leading to resolution of all symptoms in the vast majority of cases and, depending on how soon the diet is begun, can also eliminate the heightened risk of osteoporosis and intestinal cancer [109].

The work of Dicke in 1950 has proved that wheat and rye damage the small-intestinal mucosa of patients with coeliac disease. The injurious constituent of wheat in patients with coeliac disease is α -gliadin in the prolamin fraction of wheat gluten [32]. Oats do not contain gliadin but its counterpart avenin [12]. In wheat, rye, and barley, prolamins constitute 40–50, 30–50, and 35–45% of total proteins, respectively. However, in case of oats, prolamins constitute only 10–15% of total proteins and sixty grams of oats is estimated to contain 1.2 g of avenin [20].

Generally, it is recommended that coeliac patients should avoid wheat, rye, and barley. The removal of gluten from their diet results in epithelial healing and the gradual reformation of intestinal villi. Although the inclusion of oats in the gluten free diet was controversial until 1996, several later studies indicate that oats are not unsafe for those either with coeliac disease or dermatitis herpetiformis. Moreover, adherence to a strict gluten-free diet is difficult and any relief of dietary restrictions, such as those on oats, can make

the diet more acceptable to patients [65]. Oats improve the nutritional value of the gluten-free diet without any negative effects on nutritional status and are appreciated by the patients. Therefore, oats can help coeliac patients following a strict gluten-free diet [105].

The safety of oats in individuals with coeliac disease has been extensively investigated. Clinical evidence confirms that consumption of pure, uncontaminated oats is safe up to 50–70 g/day by adults and 20 to 25 g/day for children. Studies looking at the consumption of oats over 5 years have confirmed their safety. Such studies have increased the possibility of adding oats to a gluten-free diet, as this would permit a wider choice of foods for individuals with coeliac disease and provide an additional source of carbohydrates, proteins and fiber. However, some people with coeliac disease show no improvement on the gluten-free diet, the condition called unresponsive coeliac disease. The most common reason for poor response is that small amount of gluten fraction is still present in the diet. Pure oats are being grown and produced following good agricultural practices to minimize the presence of other cereals. This helps to make sure that gluten from other grains is not mixed with the pure oats [42].

The effects of gluten-free diets with and without oats (50–70 g/day) in adults with coeliac disease have been studied [53]. The oat and control groups did not differ significantly in nutritional status, symptoms, or in vitro testing. Patients in remission, regardless of diet, did not have worsening effect on the duodenal villi. All the patients with new diagnoses were in remission at 1 year, except for one in the control group. It is likely that the results apply to children as well, because the toxicity of oats in children would be difficult to test owing to the extended time span required for relapse. Therefore, many patients with coeliac disease can include moderate amounts of oats to their gluten-free diets without harmful effects as supported by other studies as well [46, 52, 90, 103].

In a long-term study, 32 children with coeliac disease were either challenged with oats or with gluten. When relapse was evident after gluten challenge, a gluten-free diet including oats was started to this group. At the end, in coeliac children in remission, oats had no detrimental effect on intestinal histology or serology. In contrast, the gluten challenge group relapsed after 3–12 months. Complete recovery from the disease was accomplished in all relapsed and newly detected patients on an oat-containing gluten-free diet. After the trial, 86% of the children preferred to consume oats and all of them remained in remission [47]. Similar results have been described previously in the adults [52].

Some in vitro studies also indicate lack of oats toxicity in coeliac disease patients. It is well documented that the wheat protein gliadin triggers inflammation in coeliac patients. However, the potential toxicity of avenin, the corresponding protein in oats has been evaluated and the results showed that the immunogenic sequences present in gliadin are missing in avenin [68].

Picarelli et al. [84] cultured intestinal biopsy tissue specimens from subjects with quiescent celiac disease to determine if oats were a source of toxicity. Samples of duodenal mucosa that were obtained by biopsy were cultured in media containing immunologically active components of wheat (gliadin) or oats (avenin). Tissue cultures produced celiac-specific antibodies only to gliadin, whereas there was no antibody response to avenin or its most toxic fraction. These results suggest that oats can be included safely in a gluten-free diet for celiac disease.

Kilmartin et al. [69] examined the immune response of celiac mucosal T-cell lines to prolamin fractions from wheat, rye, barley, and oats. They isolated mucosal T-cells from coeliac patients and investigated their interaction with the prolamin fraction of the four cereals. The T-cell lines demonstrated immunoreactivity to protein fractions from the four related cereals. However, despite oats stimulation of T cell lines, it did not activate mucosal lesion in celiac patients. The treatment with tTG enhanced the response to gliadin, secalin, and hordein, but the enzyme caused little or no enhancement of avenin responsiveness. The lower number of proline residues in avenin makes the protein less suitable substrate for tTG, and that the binding of avenin to HLA-DQ2 is less efficient than the r prolamins of other cereals.

A few studies have been performed to investigate the nutritional quality of the gluten free diet. These studies reported that the gluten free diet could be deficient in fiber, vitamins and minerals, especially for women [43]. Inclusion of oats in the gluten free diet is advantageous, since oats are a good source of dietary fiber and of several vitamins and minerals. Nevertheless, there could be some negative nutritional impacts in relation to its high intake, as insoluble dietary fiber may interact with needed mineral elements resulting their lower bioavailability [85, 93, 112] and increased risk of mineral deficiencies [33, 88]. The effects of complex carbohy-

drates on the absorption of minerals have been reviewed in an array of foods [19].

The decreased absorption of minerals is not only related to ion binding capacity of fiber components but some other factors such as intestinal transit time, degree of bacterial fiber degradation in gut and potential for the absorption of minerals in the large intestine may contribute in this progression. Moreover, impaired mineral availability may not be entirely due to fiber but phytate in unrefined foods may also be a causative factor [16, 97]. During food processing and digestion, phytate may be degraded to inositol phosphates with a lower phosphorylation degree thus reducing the adverse effect of phytic acid on mineral absorption. However, it should be considered that both, dietary fiber and phytic acid are present in fiber-rich diets thus it is rather difficult to separate the effects of fiber and phytate in the binding of essential polyvalent metallic ions [95, 96]. While oats are considered to have low phytase activity, heat treatment of oats aimed to protect against fat oxidation further decreases the activity of enzyme [40].

Conclusion

Various strategies are used to combat diseases and potential health risks. In addition to pharmaceutical approach, diet based strategies are also considered suitable to prevent various disorders. In the developing countries, increased cost of medication and their side effects are of great concern to general public; opening new channels of pharmacological investigations focusing on natural medication and diverting human trends toward natural cure. Manipulation of different foods can be helpful to control different diseases. Oat grains are good source of B complex vitamins, protein, fat, minerals, and heart healthy soluble- fiber β -glucan. Moreover it is also useful for the control of diabetes and lipid profile. The incorporation of oat in daily diet is not only important from the nutrition standpoint but also for its therapeutic potential. However, the health claims associated with oats are achieved only when oat products are consumed regularly for longer time and the amount in the diet overcomes certain threshold values set for β -glucan.

References

1. AACC (2001) The definition of dietary fibre. *Cereal Foods World* 46:112
2. Anderson JW, Chen WL (1986) Cholesterol-lowering properties of oat products. In Webster FH (ed) *Oats, Chemistry and Technology*, Am Assoc Cereal Chem, St Paul, MN, pp 309–333
3. Aman P, Rimsten L, Andersson R (2004) Molecular weight distribution of β -glucan in oat-based foods. *Cereal Chem* 81:356–360
4. Anderson JW (1995) Dietary fibre, complex carbohydrate, and coronary artery disease. *Can J Cardiol* 11(Suppl G):55G–62G
5. Anderson JW, Gilinsky NH, Deakins DA, Smith SF, O'Neal DS, Dillon DW, Oeltgen PR (1991) Lipid responses of hypercholesterolemic men to oat-bran and wheat-bran intake. *Am J Clin Nutr* 54:678–683
6. Anderson JW, Story L, Siding B, Chen WJ, Petro MS, Story J (1984) Hypocholesterolemic effects of oat-bran or bean intake for hypercholesterolemic men. *Am J Clin Nutr* 40:1146–1155
7. Anderson JW, Tietjen-Clark J (1986) Dietary fiber: hyperlipidemia, hypertension, and coronary heart disease. *Am J Gastroenterol* 81:907–919
8. Anonymous (1989) AACC committee adopts oat bran definition. *Cereal Chem* pp 1–24
9. Autio K, Myllymäki O, Mälikki Y (1987) Flow properties of solutions of oat β -glucans. *J Food Sci* 52:1354–1366
10. Autio K, Myllymäki O, Suortti T, Saastamoinen M, Poutanen K (1992) Physical properties of (1 \rightarrow 3) (1 \rightarrow 4)- β -D-glucan prepartes isolated from Finnish oat varieties. *Food Hydrocolloid* 5:513–522
11. Bach Knudsen KE, Jensen BB, Hansen I (1993) Digestion of polysaccharides and other major components in the small and large intestine of pigs fed on diets consisting of oat fractions rich in β -D-glucan. *British J Nutr* 70:537–556
12. Baker PG (1974) Oats and coeliac disease. *BMJ* 4:588–589
13. Beer M, Wood P, Weisz J, Fillion N (1997) Effect of cooking and storage on the amount and molecular weight of (1 \rightarrow 3)(1 \rightarrow 4)- β -D-glucan extracted from oat products by an in vitro digestion system. *Cereal Chem* 74:705–709
14. Bell LP, Hectorn KJ, Reynolds H, Hunninghake DB (1990) Cholesterol lowering effects of soluble-fiber cereals as part of a prudent diet for patients with mild to moderate hypercholesterolemia. *Am J Clin Nutr* 52:1020–1026
15. Beynen AC, Buechler KF, Van der Molen AT, Geelen MJH (1982) The effects of lactate and acetate on fatty acid and cholesterol biosynthesis by isolated rat hepatocytes. *Int J Biochem* 14:165–169
16. Bitar K, Reinhold JG (1972) Phytase and alkaline phosphatase activities in intestinal mucosae of rat, chicken, calf and man. *Biochim Biophys Acta* 268:442–452
17. Braaten TJ, Wood PJ, Scott FW, Wolynetz MS, Lowe MK, Bradley-Whyte P (1994) Oat β -glucan reduces blood cholesterol concentration in hypercholesterolemic subjects. *Eur J Clin Nutr* 48:465–474
18. Bridges SR, Anderson JW, Deakins DA, Dillon DW, Wood CL (1992) Oat bran increases serum acetate of hypercholesterolemic men. *Am J Clin Nutr* 56:455–459
19. British Nutrition Foundation (1990) *Complex carbohydrates in food. Report of British Nutrition Foundation*, Chapman and Hall, London
20. Brohult S, Sandegren E (1954) In: Neurath H, Bailey KC (eds) *Proteins: chemistry, biological activity, and methods*, Part B, vol 2. Academic Press, New York, p 487
21. Brown MS, Goldstein JL (1983) Lipoprotein receptors in the liver-I control signals for plasma cholesterol traffic. *J Clin Invest* 72:743–747
22. Byington RP, Jukema JW, Salonen JT (1995) Reduction in cardiovascular events during pravastatin therapy: pooled analysis of clinical events of the Pravastatin Atherosclerosis Intervention Program. *Circulation* 92:2419–2425
23. Campbell GL, Bedford MR (1992) Enzyme applications for mono-gastric feeds: a review. *Can J Anim Sci* 72:449–466
24. Cara L, Dubois C, Borel P, Armand M, Senfi M, Portugal H, Paull A, Bernard P, Lairon D (1992) Effects of oat bran, rice bran, wheat fiber, and wheat germ on postprandial lipemia in healthy adults. *Am J Clin Nutr* 55:81–88
25. Chen WL, Anderson JW (1986) Hypocholesterolemic effects of soluble fiber. In: Vahony GV, Kritchevsky D (eds) *Dietary fiber basic and clinical aspects*, Plenum Press, New York, pp 275–286
26. Colleoni-Sirghie M, Kovalenko IV, Briggs JL, Fulton B, White PJ (2003) Rheological and molecular properties of water soluble (1 \rightarrow 3) (1 \rightarrow 4)- β -D-glucans from high- β -glucan and traditional oat lines. *Carbohydr Polym* 52:439–447
27. Cummings JH, Pomare EW, Branch WJ, Naylor CP, Macfarlane GT (1987) Short chain fatty acids in human large intestine, portal, hepatic and venous blood. *Gut* 28:1221–1227
28. Dais PJ, Perlin AS (1982) High field, ^{13}C -NMR spectroscopy of β -D-glucan, amylopectin and glycogen. *Carbohydr Res Chem* 71:301–307
29. Dawkins NL, Nnanna IA (1995) Studies on oat gum [(1 \rightarrow 3, 1 \rightarrow 4)- β -D-glucan]: composition, molecular weight estimation and rheological properties. *Food Hydrocolloid* 9:1–7
30. De Schrijver R, Fremaut D, Verheyen A (1992) Cholesterol lowering effects and utilization of protein, lipid, fiber and energy in rats fed unprocessed and baked oat bran. *J Nutri* 122:1318–1324
31. Deguyte-Fomins L, Sontag-Strohm T, Salovaara H (2002) Oat bran fermentation by rye sourdough. *Cereal Chem* 79:345–348
32. Dicke WK, Weijers HA, van de Kamer JH (1953) Coeliac disease. II. The presence in wheat of a factor having a deleterious effect in cases of coeliac disease. *Acta Paediatr* 42:34–42
33. Donangelo CM, Eggum BO (1986) Comparative effects of wheat bran and barley husk on nutrient utilization in rats. 2. Zinc, calcium and phosphorus. *Br J Nutr* 56:269–280
34. Dongowski G, Drzikova B, Senge B, Blochwitz R, Gebhardt E, Habel A (2005) Rheological behaviour of β -glucan preparations from oat products. *Food Chem* 93:279–291
35. Doublier JL, Wood PJ (1995) Rheological properties of aqueous solutions of (1 \rightarrow 3) (1 \rightarrow 4)- β -D-glucan from oat (*Avena sativa* L.). *Cereal Chem* 72:335–340
36. Drzikova B, Dongowski G, Gebhardt E (2005) Dietary fibre-rich oat-based products affect serum lipids, microbiota, formation of short-chain fatty acids and steroids in rats. *Br J Nutr* 94(6):12–25

37. Drzikova B, Dongowski G, Gebhardt E, Habel A (2005) The composition of dietary fiber rich extrudates from oat affects bile acid binding and fermentation in vitro. *Food Chem* 90:181–192
38. Fasano A, Berti I, Gerarduzzi T, Not T, Colletti RB, Drago S, Elitsur Y, Green PH, Guandalini S, Hill ID, Pietzak M, Ventura A, Thorpe M, Kryszak D, Fornaroli F, Wasserman SS, Murray JA, Horvath K (2003) Prevalence of celiac disease in at risk and not at risk groups in the United States: a large multi center study. *Arch Intern Med* 163:286–292
39. Fleming SE, Fitch MD, Chansler MW (1989) High-fiber diets: influence on characteristics on cecal digesta including short-chain fatty acid concentrations and pH. *Am J Clin Nutr* 50:93–99
40. Frolich W, Nyman M (1988) Minerals, phytate and dietary fibre in different fractions of oat grain. *J Cereal Sci* 7:73–82
41. Glore SR, Van TD, Knehans AW, Guild M (1994) Soluble fiber and serum lipids: a literature review. *J Am Diet Assoc* 94:425–436
42. Green PHR, Jabri B (2006) Celiac Disease. *Annu Rev Med* 57:207–221
43. Grehn S, Fridell K, Lilliecreutz M, Hallert C (2001) Dietary habits of Swedish adult coeliac patients treated by a glutenfree diet for 10 years. *Scand J Nutr* 45:178–182
44. Guynn RW, Veech RL (1975) Enzymatic determination of acetate. In: Lowenstein JM (ed) *Methods of enzymology*, vol 35. Academic Press, New York, pp 302–307
45. Haack VS, Chesters JG, Vollendorf NW, Story JA, Marlett JA (1998) Increasing amounts of dietary fiber provided by foods normalizes physiologic response of the large bowel without altering calcium balance or fecal steroid excretion. *Am J Clin Nutr* 68:615–622
46. Hardman CMN, Garioch JJ, Leonard JN, Thomas HJ, Walker MM, Lortan JE, Lister A, Fry L (1997) Absence of toxicity of oats in patients with dermatitis herpetiformis. *N Engl J Med* 337:1884–1887
47. Holm K, Maki M, Vuolteenaho N, Mustalahti K, Ashorn M, Ruuska T, Kaukinen K (2006) Oats in the treatment of childhood coeliac disease: a 2-year controlled trial and a long-term clinical follow-up study. *Aliment Pharmacol Ther* 23:1463–1472
48. Idouraine A, Khan MJ, Weber CW (1996) In Vitro Binding Capacity of Wheat Bran, Rice Bran, and Oat Fiber for Ca, Mg, Cu, and Zn Alone and in Different Combinations. *J Agric Food Chem* 44:2067–2072
49. Jacobs DR, Marquart L, Slavin J, Kushi LH (1998a) Whole-grain intake and cancer: an expanded review and metaanalysis. *Nutr Cancer* 30:85–96
50. Jacobs DR, Marquart L, Slavin J, Kushi LH (1998b) Whole grain intake may reduce the risk of ischemic heart disease death in postmenopausal women: The Iowa Women's Health Study. *Am J Clin Nutr* 68:248–257
51. Jacobs DR, Meyer KA, Kushi LH, Folsom AR (1998) Whole-grain intake may reduce the risk of ischemic heart disease death in postmenopausal women: the Iowa Women's Health Study. *Am J Clin Nutr* 68:248–257
52. Janatuinen EK, Kemppainen TA, Pikkarainen PH, Holm KH, Kosma VM, Uusitupa MI, Maki M, Julkunen RJ (2000) Lack of cellular and humoral responses to oats in adults with coeliac disease. *Gut* 46:327–331
53. Janatuinen EK, Pikkarainen PH, Kemppainen TA, Kosma VM, Järvinen RM, Uusitupa MI, Julkunen RJ (1995) A comparison of diets with and without oats in adults with celiac disease. *N Engl J Med* 333(16):1033–1037
54. Jenkins AL, Jenkins DJA, Zdravkovic U, Würsch P, Vuksan V (2002) Depression of the glycemic index by high levels of beta-glucan fiber in two functional foods tested in type 2 diabetes. *Eur J Clin Nutr* 56(7):622–628
55. Jenkins DJ, Wolever TM, Vuksan V, Brighenti F, Cunnane SC, Rao AV, Jenkins AL, Buckley G, Patten R, Singer W (1989) Nibbling versus gorging: metabolic advantages of increased meal frequency. *N Engl J Med* 321(14):929–934
56. Johansen HN, Bach Knudsen KE, Wood PJ, Fulcher RG (1997) Physicochemical properties and the degradation of oat bran polysaccharides in the gut of pigs. *J Sci Food Agr* 73:81–92
57. Judith H, Daniel JS, Behall KM (1995) Diets containing soluble oat extracts improve glucose and insulin responses of moderately hypercholesterolemic men and women. *Am J Clin Nutr* 61:379–384
58. Kahlon TS, Chow FI (1997) Hypocholesterolemic effects of oat, rice, and barley dietary fibers and fractions. *Cereal Food World* 42:86–92
59. Kahlon TS, Chow FI, Knuckles BE, Chiu MM (1993) Cholesterol lowering effects in hamsters of β -glucan enriched barley fraction, dehulled whole barley, rice bran, and oat bran and their combinations. *Cereal Chem* 70:435–440
60. Kahlon TS, Chow FI, Sayre RN, Betschart AA (1992a) Cholesterol lowering in hamsters fed rice bran at various levels, defatted rice bran and rice bran oil. *J Nutr* 122:513–519
61. Kahlon TS, Chow FI, Wood DF (1999) Cholesterol Response and Foam Cell Formation in Hamsters Fed Rice Bran, Oat Bran, and Cellulose + Soy Protein Diets With or Without Added Vitamin E. *Cereal Chem* 76(5):772–776
62. Kahlon TS, Saunders RM, Chow FI, Chiu MM, Betschart AA (1990) Influence of rice bran, oat bran, and wheat bran on cholesterol and triglycerides in hamsters. *Cereal Chem* 67:439–443
63. Kahlon TS, Saunders RM, Sayre RN, Chow FI, Chiu MM, Betschart AA (1992b) Cholesterol-lowering effects of rice bran and rice bran oil fractions in hypercholesterolemic hamsters. *Cereal Chem* 69:485–489
64. Kannel WB, Castelli WD, Gordon T, McNamara PM (1971) Serum cholesterol, lipoproteins, and risk of coronary artery disease. The Framingham Study. *Ann Intern Med* 74:1–12
65. Kasarda DD (2000) Celiac disease. In: Kiple KF, Ormelas C (eds) *The Cambridge World History of Food*, vol 1. Cambridge University Press, Cambridge, pp 1008–1022
66. Kerckhoffs D, Hornstra G, Mensink R (2003) Cholesterol-lowering effect of β -glucan from oat bran in mildly hypercholesterolemic subjects may decrease when β -glucan is incorporated into bread and cookies. *Am J Clinical Nutr* 78:221–227
67. Kestin M, Moss R, Clifton PM, Nestel PJ (1990) Comparative effects of three cereal brans on plasma lipids, blood pressure, and glucose metabolism in mildly hypercholesterolemic men. *Am J Clin Nutr* 52:661–666
68. Kilmartin C, Lynch S, Abuzakouk M, Wieser H, Feighery C (2003) Avenin fails to induce a Th1 response in coeliac tissue following in vitro culture. *Gut* 52(1):47–52
69. Kilmartin C, Wieser H, Abuzakouk M, Kelly J, Jackson J, Feighery C (2006) Intestinal T cell responses to cereal proteins in celiac disease. *Dig Dis Sci* 51(1):202–209
70. Kirby RW, Anderson JW, Sieling B, Rees D, Chen WJL, Miller RE, Kay RM (1981) Oat-bran intake selectively lowers serum low-density lipoprotein cholesterol concentrations of hypercholesterolemic men. *Am J Clin Nutr* 34:824–829
71. Kirk RS, Sawyer R (1999) Pearson's composition and analysis of foods, 9th edn. Addison-Wesley Longman Inc., Harlow, England, p 285

72. Kris-Etherton PM, Krummel D, Russell ME, Dreon D, Mackey S, Borchers J, Wood PD (1988) The effect of diet on plasma lipids, lipoproteins, and coronary heart disease. *J Am Diet Assoc* 88:1373–1400
73. Kupper C (2005) Dietary guidelines and implementation for celiac disease. *Gastroenterology* 128(4 Suppl 1):S121–S127
74. Lia Å, Hallmans G, Sandberg AS, Sundberg B, Åman P, Andersson H (1995) Oat β -glucan increases bile acid excretion and a fiber-rich barley fraction increases cholesterol excretion in ileostomy subjects. *Am J Clin Nutr* 62:1245–1251
75. Lyly M, Salmenkallio-Marttila M, Suortti T, Autio K, Poutanen K, Lahteenmaki L (2003) Influence of oat β -glucan preparations on the perception of mouth feel and rheological properties in beverage prototypes. *Cereal Chem* 80:536–541
76. Magness JR, Markle GM, Compton CC (1971) Food and feed crops of the United States, 1st edn. Interregional Research Project IR-4, New Jersey. Bul 828
77. Mahley RW (1982) Atherogenic hyperlipoproteinemia. The cellular and molecular biology of plasma lipoproteins altered by dietary fat and cholesterol. *Med Clin North Am* 66:375–401
78. Marlett JA (1993) Comparisons of dietary fiber and selected nutrient compositions of oat and other grain fractions. In: Wood PJ (ed) Oat bran. American Association of Cereal Chemists, St. Paul, MN, pp 49–82
79. McIntosh GH, Whyte J, McArthur R, Nestel PJ (1991) Barley and wheat foods influence on plasma cholesterol concentrations in hypercholesterolemic men. *Am J Clin Nutr* 53:1205–1209
80. Miller SS, Fulcher RG, Sen A, Arnason JT (1995) Oat endosperm cell walls: I. Isolation, composition, and comparison with other tissues. *Cereal Chem* 72:421–427
81. Mälikki Y (2001) Physical properties of dietary fiber as keys to physiological functions. *Cereal Food World* 46:196–199
82. Mälikki Y, Virtanen E (2001) Gastrointestinal effects of oat bran and oat gum a review. *Lebensmittel-Wissenschaft Technol* 34:337–347
83. Parrish FW, Perlin AS, Reese ET (1960) Selective enzymolysis of poly- β -D-glucan and structure of the polymers. *Can J Chem* 38:2094–2104
84. Picarelli A, Di Tola M, Sabbatella L, Gabrielli F, Di Cello T, Anania MC, Mastracchio A, Silano M, De Vincenzi M (2001) Immunologic evidence of no harmful effect of oats in celiac disease. *Am J Clin Nutr* 74: 137–140
85. Platt SR, Clydesdale FM (1985) Binding of iron by Lignin in the presence of various concentrations of Calcium, Magnesium, and Zinc. *J Food Sci* 50(5):1322–1326
86. Qureschi AA, Burger WC, Peterson DM, Elson C (1985) Suppression of cholesterologenesis by plant constituents: review of Wisconsin contribution to NC-167. *Lipids* 20:817–824
87. Redard CL, Davis PA, Schneeman BO (1990) Dietary fiber and gender: effect on postprandial lipemia. *Am J Clin Nutr* 52:837–845
88. Reinhold JG, Faradji B, Abadi P, Ismail-Beige F (1976) Decreased absorption of calcium, magnesium, zinc and phosphorus by humans due to increased fiber and phosphorus consumption as wheat bread. *J Nutr* 106:493–503
89. Remesy C, Demigne C, Chartier F (1980) Origin and utilization of fatty acids in the rat. *Reprod Nutr Dev* 30:1339–1349
90. Reunala T, Collin P, Holm K, Pikkarainen P, Miettinen A, Vuolteenaho N, Mäki M (1998) Tolerance to oats in dermatitis herpetiformis. *Gut* 43:490–493
91. Ripsin CM, Keenan JM, Jacobs DR, Elmer PJ, Welch RR, Van Horn L, Liu K, Turnbull WH, Thye FW, Kestin M (1992) Oat products and lipid lowering: a meta-analysis. *JAMA* 267:3317–3325
92. Robertson JA, Majsak-Newman G, Ring SG (1997) Release of mixed linkage (1 \rightarrow 3), (1 \rightarrow 4) b-D-glucans from barley by protease activity and effects on ileal effluents. *Intern J Biol Macromol* 21:57–60
93. Rossander-Hulthen L, Glerup A, Hallberg L (1990) Inhibitory effect of oat products on non-haem iron absorption in man. *Eur J Clin Nutr* 44:783–791
94. Sacks FM, Pfeffer MA, Moye LA, Rouleau JL, Rutherford JD, Cole TG, Brown L, Warnica JW, Arnold JM, Wun CC, Davis BR, Braunwald E (1996) The effect of pravastatin on coronary events after myocardial infarction in patients with average cholesterol levels. *N Engl J Med* 335:1001–1009
95. Sandberg AS, Andersson H, Carleeon NG, Sandstram B (1987) Degradation products on bran phytate formed during digestion in the human small intestine: effect of extrusion cooking on digestibility. *J Nutr* 117:2061–2065
96. Sandberg AS, Carleeon NG, Svanberg U (1989) In vitro studies of inositol tri-, tetra-, penta- and hexaphosphates as potential iron absorption inhibitors. In: Southgate DAT, Johnson I, Fenwick GR (eds) Nutrient availability. Royal Society of Chemistry, Cambridge, pp 158–160
97. Sandstorm B, Almgren A, Kivistd B, Cederblad A (1987) Zinc absorption in humans from meals based on rye, barley, oatmeal, triticale and whole wheat. *J Nutr* 117:1898–1902
98. Saunders RM (1985) Rice bran: Composition and potential food uses. *Food Rev Int* 1:465–495
99. Seibert SE (1987) Oat bran as a source of soluble dietary fiber. *Cereal Food World* 32:552–553
100. Shan L, Molberg O, Parrot I, Hausch F, Filiz F, Gray GM, Sollid LM, Khosla C (2002) Structural basis for gluten intolerance in celiac sprue. *Science* 297:2275–2279
101. Slavin J, Marquart L, Jacobs D Jr (2000) Consumption of whole grain foods and decreased risk of cancer: Proposed mechanisms. *Cereal Food World* 45:54–58
102. Small E (1999) New crops for Canadian agriculture. In: Janick J (ed) Perspectives on new crops and new uses. ASHS Press, Alexandria, VA, pp 15–52
103. Srinivasan U, Leonard N, Jones E, Kasarda DD, Weir DG, O'Farrelly C, Feighery C (1996) Absence of oats toxicity in adult coeliac disease. *BMJ* 313:1300–1301
104. Storer GB, Trimble RP, Illman RI (1983) Effects of dietary oat bran and diabetes on plasma and caecal volatile fatty acids in the rat. *Nutr Res* 3:519–526
105. Storsrud S, Hulthen LR, Lenner RA (2003) Beneficial effects of oats in the gluten-free diet of adults with special reference to nutrient status, symptoms and subjective experiences. *Brit J Nutr* 90:101–107
106. Sundberg B, Wood P, Lia Å, Andersson H, Sandberg AS, Hallmans G (1996) Mixed-linked β -glucan from breads of different cereals is partly degraded in the human ileostomy model 1–3. *Am J Clin Nutr* 64:878–885
107. Tapola N, Karvonen H, Niskanen L, Mikola M, Sarkkinen E (2005) Glycemic responses of oat bran products in type 2 diabetic patients. *Nutr Metab Cardiovas* 15(4):255–261
108. Thompson LU (1994) Antioxidants and hormone-mediated health benefits of whole grains. *Crit Rev Food Sci Nutr* 34:473–497
109. Treem W (2004) Emerging concepts in celiac disease. *Curr Opin Pediatr* 16(5):552–559

110. Trowell HC, Burkitt DP (1981) Western diseases: their emergence and prevention. Harvard University Press, Cambridge, pp 227–267
111. Truswell AS (1995) Dietary fibre and plasma lipids. *Eur J Clin Nutr* 49(suppl):5105–5109
112. Walker A (1985) Mineral metabolism. In: Trowell H, Burkitt D, Heaton K (eds) Dietary fibre, fibre-depleted food and disease. Academic Press London, pp 361–375
113. Welch R, Brown J, Leggett J (2000) Interspecific and intraspecific variation in grain and groat characteristics of wild oat (*Avena*) species: Very high groat (1–3), (1–4)- β -D-glucan in an *Avena atlantica* genotype. *J Cereal Sci* 31:273–279
114. Whole Grains Bureau. History of Whole Grains. http://www.wholegrainsbureau.ca/about_wg/history_of_wg.html. Accessed 9 April 2007
115. Wikstrom K, Lindahl L, Andersson R, Westerlund E (1994) Rheological studies of water-soluble (1 \rightarrow 3) (1 \rightarrow 4)- β -D-glucans from milling fractions of oat. *J Food Sci* 59:1077–1080
116. Wisker E, Feldheim W, Pomeranz Y, Meuser F (1985) Dietary fiber in cereals. In: Pomeranz Y (ed) *Advances in Cereal Science and Technology*, Vol 7. Am Assoc Cereal Chem, St. Paul, MN, pp 169–238
117. Wood PJ (1984) Physicochemical properties and technological and nutritional significance of cereal β -glucans. In: Rasper VF (ed) *Cereal Polysaccharides in Technology and Nutrition*. Am Assoc Cereal Chem, St Paul, MN, pp 52–57
118. Wood PJ (1991) Oat β -glucan physicochemical properties and physiological effects. *Trends Food Sci Technol* 2:311–314
119. Wood PJ (1993) Physicochemical characteristics and physiological properties of oat (1 \rightarrow 3) (1 \rightarrow 4)- β -D-glucan. In: Wood PJ (ed) *Oat bran*. AACC Inc, St. Paul, pp 49–82
120. Wood PJ, Braaten JT, Scott FW, Riedel D, Poste LMJ (1990) Comparisons of viscous properties of oat and guar gum and the effects of these and oat bran on glycemic index. *J Agric Food Chem* 38:753–757
121. Wood PJ, Braaten JA, Scott FD, Riedel KD, Wolynetz MS, Collins MW (1994) Effect of dose and modification of viscous oat gum on plasma glucose and insulin following an oral glucose load. *British J Nutrition* 72:731–743
122. Wright RW, Anderson JW, Bridges SR (1990) Propionate inhibits hepatic lipid syntheses. *Proc Soc Exp Biol Med* 195:26–29
123. Zhang JX, Hallmans G, Andersson H (1992) Effect of oat bran on plasma cholesterol and bile acid excretion in nine subjects with ileostomies. *Am J Clin Nutr* 56:99–105